

SPICED – Symposium

Survival, detection and toxinogenic potential of *B. cereus* group species in spices and herbs

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B. cereus group (= B. cereus sensu lato)

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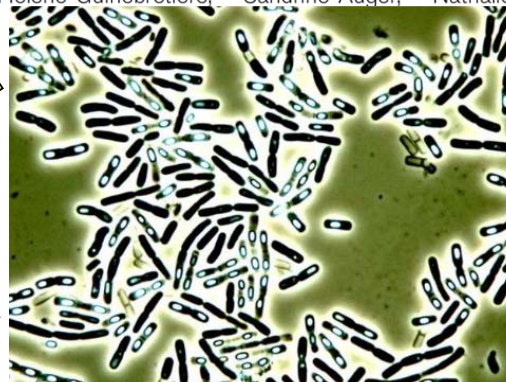
https://commons.wikimedia.org/wiki/File:Bucatini_%28amatriciana_rossa%29.jpg

B. cereus

B. cytotoxicus

Bacillus cytotoxicus sp. nov. is a novel thermotolerant species of the *Bacillus cereus* Group occasionally associated with food poisoning

Marie-Hélène Guinebretière,^{1,2} Sandrine Auger,^{3,4} Nathalie Galleron,^{3,4}



[micbio.wzw.tum.de/cms/docs/Stammsammlung/Foto Bibliothek Bacillus.pdf](http://micbio.wzw.tum.de/cms/docs/Stammsammlung/Foto%20Bibliothek%20Bacillus.pdf)

B. anthracis



<http://phil.cdc.gov/phil/details.asp>



B. thuringiensis

Wagner, A. (2015)

B. mycoides and pseudomycoides



[micbio.wzw.tum.de/cms/docs/Stammsammlung/Foto Bibliothek Bacillus.pdf](http://micbio.wzw.tum.de/cms/docs/Stammsammlung/Foto%20Bibliothek%20Bacillus.pdf)

B. weihenstephanensis

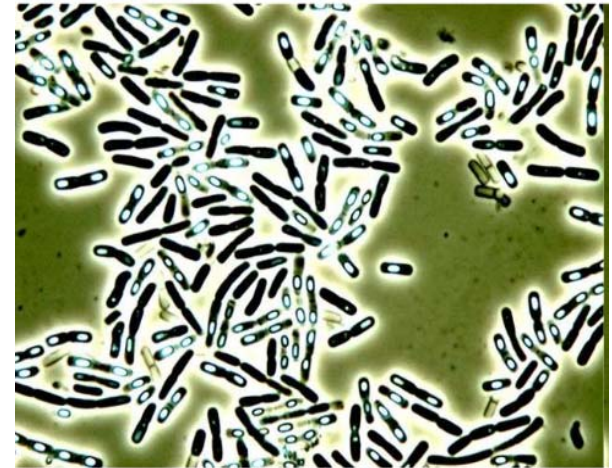


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Bacillus (B.) cereus

- Gram-positive, motile, endospore forming rod
- facultative anaerobic
- optimal growth at 30 - 37 °C
- growth limitations: 4 °C, minimum pH 4.1, lowest aw 0.92
- heat resistant spores (survive pasteurization and cooking for several minutes)
- prevention of growth after cooking at $T < 8\text{ °C}$ (optimal $< 4\text{ °C}$) or $> 63\text{ °C}$
- forms biofilms
- toxin producer
- ubiquitous in soil
- detectable in many kinds of food



micbio.wzw.tum.de/cms/docs/Stammsammlung/
Foto Bibliothek Bacillus.pdf

EFSA, 2005



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Outbreaks

Spice	Year	Country	Cases	Reference
Spicemix	2007	France	146	EFSA (2009, 2013)
Paprika	2009	Denmark	48	FAO und WHO (2014)
Curry	2009	Belgium	7	EFSA (2011)
White pepper	2010	Denmark	112	EFSA (2011, 2013)
Cinnamon	2011	Denmark	30	FAO und WHO (2014)
Turmeric	2011	Finland	19	EFSA (2013a)
Turmeric	2011	Finland	4	EFSA (2013a)
Cumin	2011	Finland	3	EFSA (2013a)
Black pepper	2011	Denmark	52	EFSA (2013a)

Food borne disease outbreaks from 1973 to 2012 in Europe associated to spices contaminated with *B. cereus*; modified according to Mader & Schaarschmidt, 2016



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Toxins

Enterotoxins

- nonhemolytic enterotoxin (Nhe)
- hemolysin BL (HBL)
- cytotoxin K (CytK)

- infective dose: $>10^5$ - 10^7 cells/spores consumed
- production in the gut
- symptoms: abdominal pain and diarrhoea
- incubation time: 8 - 16 h
- duration: 12 - 24 h
- associated to meat, vegetables, milk products

Emetic toxin (heat stable)

- cereulide

- ,effective' dose: $\sim 8 \mu\text{g}/\text{kg}$ body weight (also $0.02 - 2 \mu\text{g}/\text{kg}$)
- production in food ($10^5 - 10^8$ cells)
- symptoms: nausea and vomiting
- incubation time: 1 - 5 h
- duration 6 - 24 h
- associated to rice and pasta

Blackburn, C. de W., 2009
Ceuppens et al., 2011



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Detection – Cultural

ISO

- ❖ **7932:2005-03** Horizontal method for the enumeration of presumptive *Bacillus cereus*
- ❖ **21871:2006-04** Horizontal method for the determination of low numbers of presumptive *Bacillus cereus* (MPN)



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Detection – Cultural

Brilliance *B. cereus* Agar



PEMBA



Bacara



MYP



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Detection – Cultural

ISO

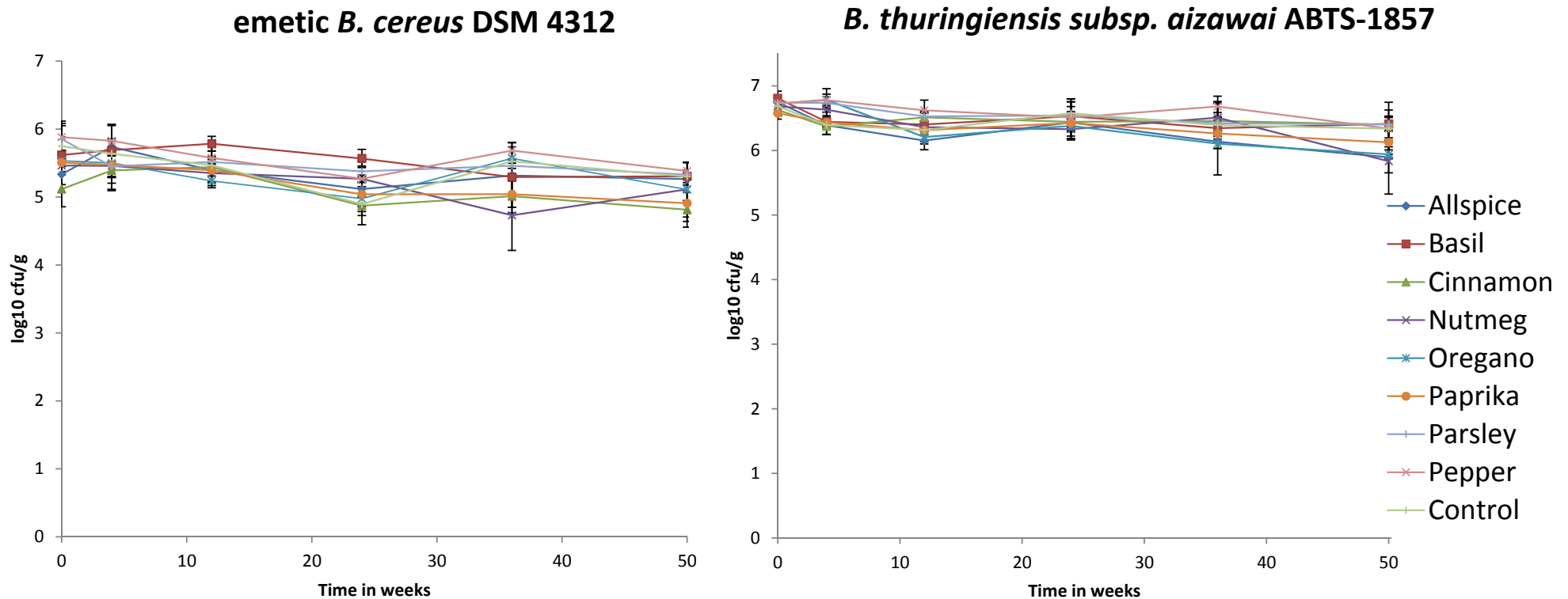
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Survival

- spore suspension air dried on 0.5 g sand + 4.5 g spice (3 parallels)
- final concentration 10^5 to 10^6 cfu/g, stored in the dark at 23 ± 1 °C.
- mean reduction rates (\log_{10}) of 0.39 ± 0.15 (*B.c.*) and 0.51 ± 0.23 (*B.t.*) after 50 weeks



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Characterization of *B. cereus* group isolates

- presumptive *B. cereus* in allspice (10^3 cfu/g), oregano, parsley (10^2 cfu/g), basil (~ 20 cfu/g), paprika and pepper (< 20 cfu/g)
- 59 isolates confirmed as *B. cereus* s.l., thereof 4 *B. weihenstephanensis*, 2 *B. mycooides* and 1 *B. thuringiensis* strain(s)
- several species in one sample
- all isolates carried toxin genes, except *B. mycooides*
- toxin gene combinations: *nheA*, *hblD* (27x); *nheA*, *hblD*, *cytK* (29x); *nheA*, *cytK*, *ces* (1x) → one emetic strain
- toxin production reflects genetic configuration (confirmed for Nhe, Hbl, cereulide)



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Multiplex real-time-PCR for *B. cereus* group species

- Testing of isolates

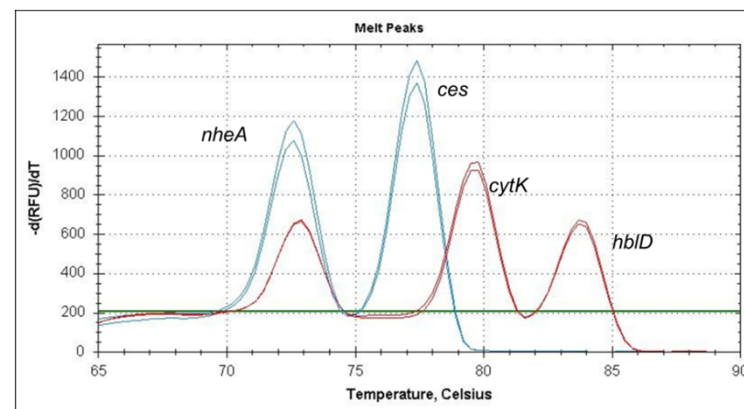
Species (Oliwa-Stasiak, K., et al., 2011; Wielinga, P.R., et al., 2011)

- *B. cereus* / *B. anthracis* / *B. thuringiensis*: *motB* gene (MotB_1)*
- *B. weihenstaphanensis*: *motB* gene (MotB_2)*
- *B. pseudomycooides*: *bpm* gene (Bpm_1)
- *cry1* positive *B. thuringiensis* strains: *cry1* plasmid gene (Tqpro_Bt)
- *B. anthracis* : lambda pro-phage type 3 sequence (Tqpro_PL3)

* *B. mycooides* with rhizoid growth

Toxin genes (Wehrle, E., et al., 2010)

HRM multiplex-PCR; for *cytK*,
nheA, *hblD* und *ces*



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Multiplex real-time-PCR for *B. cereus* group species

- Testing of spices and herbs

Species (Oliwa-Stasiak, K., et al., 2011; Wielinga, P.R., et al., 2011)

- *B. cereus* / *B. anthracis* / *B. thuringiensis*: *motB* gene (MotB_1)*
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- *B. pseudomycooides*: *bpm* gene (Bpm_1)
- *cry1* positive *B. thuringiensis* strains: *cry1* plasmid gene (Tqpro_Bt)
- Internal amplification control: pUC18 fragment (Tm_pUC18)

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DNA extraction from spores in PBS buffer

	LOD ^a (cfu/ml)	Average C _q at LOD (<i>motB</i>)
DNeasy Blood & Tissue Kit	1.4 x 10 ²	34.57 ± 0.74
MasterPure Gram Positive DNA Kit	1.4 x 10 ³	35.00 ± 1.36
Invisorb Spin Plant Mini Kit	1.4 x 10 ³	35.09 ± 1.20
Ultraclean DNA Isolation Kit	1.4 x 10 ⁴	36.51 ± 1.11
PowerSoil DNA Isolation Kit	1.4 x 10 ⁴	35.44 ± 1.25

^a Limit of detection: the lowest concentration of spores/ml (*B. cereus* DSM 4312) for which six out of six reactions (three DNA extractions and duplicate PCR reactions) gave a positive result for the *motB* target



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DNA extraction from spores in spices and herbs

		Inoculum (cfu/g) at LOD ^a	Average C _q at LOD (<i>motB</i>)
Blood and tissue	Allspice	3.3 x 10 ⁶	-
	Oregano	3.3 x 10 ⁶	34.14 ± 1.95
	Paprika	3.3 x 10 ⁵	34.43 ± 0.88
	Pepper	3.3 x 10 ⁵	35.97 ± 1.24
CTAB method	Allspice	6.2 x 10 ⁶	35.87 ± 0.89
	Oregano	6.2 x 10 ²	36.25 ± 1.29
	Paprika	6.2 x 10 ³	34.02 ± 1.38
	Pepper	6.2 x 10 ²	38.22 ± 1.18

^a Limit of detection: the lowest concentration of spores/g (*B. cereus* DSM 4312) for which twelve out of twelve reactions (three repetitions of duplicate DNA extraction and duplicate PCR) gave a positive result for the *motB* target.

CTAB (cetyltrimethylammonium bromide) method is based on ISO 21571:2013-08 and modifications according to Minarovicova et al., in press.



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Summary

- Low numbers of *B. cereus* in spices and herbs are normal → temperature regimes of dishes are crucial
- Different *B. cereus* group species with varying toxinogenic potential occur in spices and herbs
- Spores persist in spices and herbs
- Detection of *B. cereus* group species is possible at LODs of 10^2 to 10^3 spores/g with CTAB method and real-time PCR



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Thanks for your attention. Questions and comments?

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