Optimized surgical techniques and postoperative care

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Optimized surgical techniques and postoperative care

Surgery

- Optimized anaesthesia
- Pre-emptive analgesia
- Anti-infective prophylaxis
- Experienced personnel
- Surgical skills per se

Postoperative care

Pain alleviation in laboratory mice

- Pain killers

Supportive measures

- warmth, fluid, housing conditions, ...

Humane endpoints



The laboratory mouse



... represents probably ≥70% of laboratory animals used in biomedical research and testing

... thousands of genetically modified lines/models available

Examples for the need of surgery in research Organ transplantation

- liver
- Iung
- stem cells

Open chest surgery

myocardial infarction

Stroke

Physiology

. . .



Agenda: Pain alleviation in laboratory mice

Problems

- detection of pain in mice
- treating pain in mice

Example: Use of a minor surgery model for assessing (1) postsurgical pain and (2) the efficacy of the pain relief regimen Read-out:

- Clinicial investigation: cage-side observations, body weight, food consumption
- Telemetry: Heart rate, heart rate variability
- Natural behaviors: Nest building, burrowing

Where are we now?

Next steps



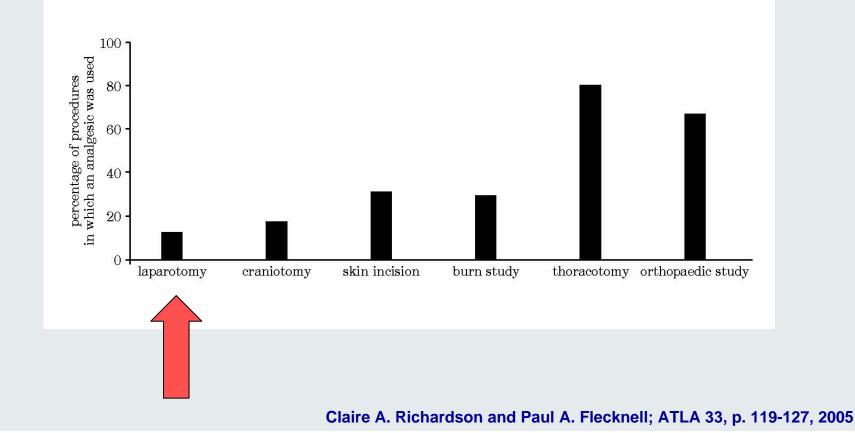
Detection of postoperative pain in mice

Examples (video recordings): No effect of pain killers visible by cage-side observations of typical clinical signs



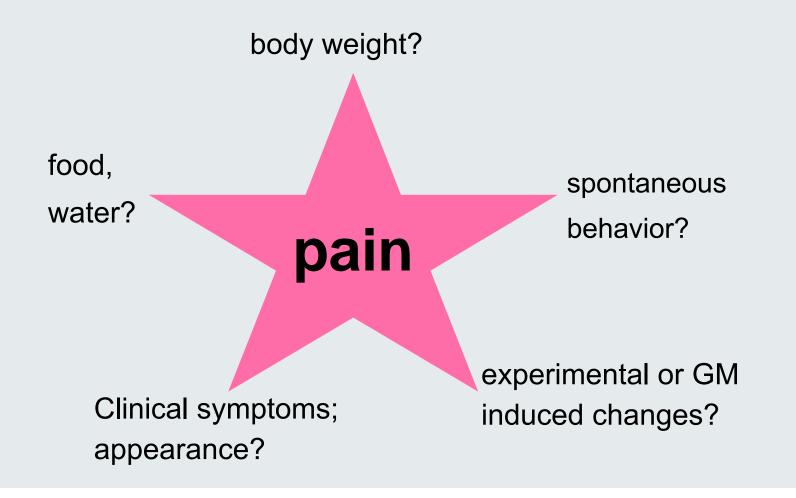
Pain treatment







Detecting ongoing pain by its effects on ...





Clinical signs

Appearance

- fur
 - rough, soiled, hair loss, piloerection
- body
 - weight loss
 - decreased food intake
 - sunken abdomen
 - hunched posture
 - sunken eyes
 - dehydration

Behavior

- aggressive
 - biting response; automutilation
- apathy
 - unresponsive; isolation from the cage mates
- increased activity
 - restlessness; sudden running movements
- prolonged times for sleeping or resting

Constraint

= fear, distress, suffering, pain, disease, ...



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Food intake after surgery

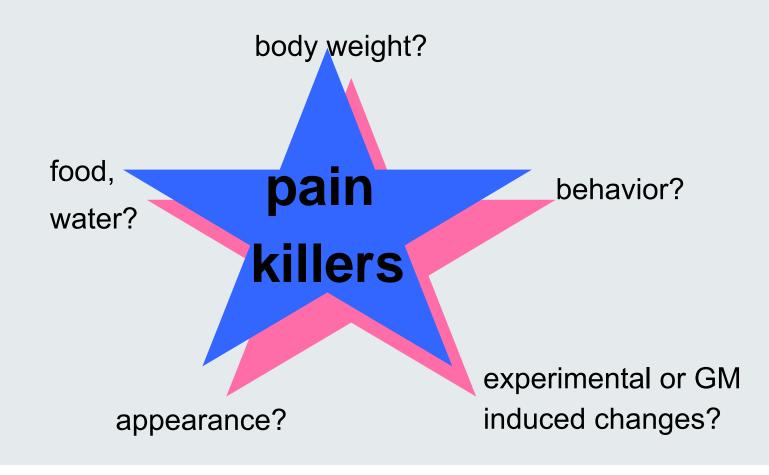
Decrease of **food consumption** after surgery Alteration of body weight, water intake, wheel runing activity Adamson 2010, Bourque 2010, Arras 2007, Bomzon 2006

Side-effects

Example (picture): Pica behaviour in the rat from Buprenorphine (Opioid) \rightarrow high dosage?



Detecting ongoing pain by its effects on ...





Obstacles in pain relief regimens

Table 3: Suggested analgesic dose rates forlaboratory rats and mice

| | Suggested analgesic dose rate | | | |
|---------------|-------------------------------------|-------------------------------|--|--|
| Analgesic | Rat | Mouse | | |
| Buprenorphine | 0.01–0.05mg/kg s.c., 6–12 hourly | 0.1mg/kg s.c., 6–12 hourly | | |
| Butorphanol | 2.0mg/kg s.c., 4 hourly | 2.0mg/kg s.c., 4 hourly | | |
| Morphine | 2–5mg/kg s.c., 4 hourly | 2–5mg/kg s.c., 4 hourly | | |

Table from Claire A. Richardson & Paul A. Flecknell, ATLA 33, p. 119-127, 2005

| Pethidine | 10–20mg/kg i.m., s.c., 2–3 hourly | 10–20mg/kg i.m., s.c., 2–3 hourly | |
|------------|--|--------------------------------------|--|
| Carprofen | 5mg/kg s.c., ? daily | 10mg/kg s.c., ? daily | |
| Meloxicam | 1–2mg/kg s.c. or 4mg/kg per os ? daily | 5mg/kg s.c. ? daily | |
| Ketoprofen | 5mg/kg s.c. ? daily | 10mg/kg s.c. ? daily | |



Pain alleviation in mice

Opioid e.g. Burpenorphine

NSAID

e.g.

Carprofen, Meloxicam, Metamizol, ...

0.1 mg/kg BW, s.c., 2/die

5 mg/kg BW, s.c., 1-2/die ?



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Problems of pain alleviation

Difficulties in detecting pain in the mouse

Side-effects of analgesics \rightarrow sometimes masking symptoms

No control of success \rightarrow efficacy of pain treatment remains questionable

Regimens (agents, dosages, application intervals) mostly rely not on systematic studies or scientific investigations

Knowledge about efficient pain treatment in mice:
→ Evidence-based regimens
= agents, dosages and intervals
→ Animal-friendly application routes



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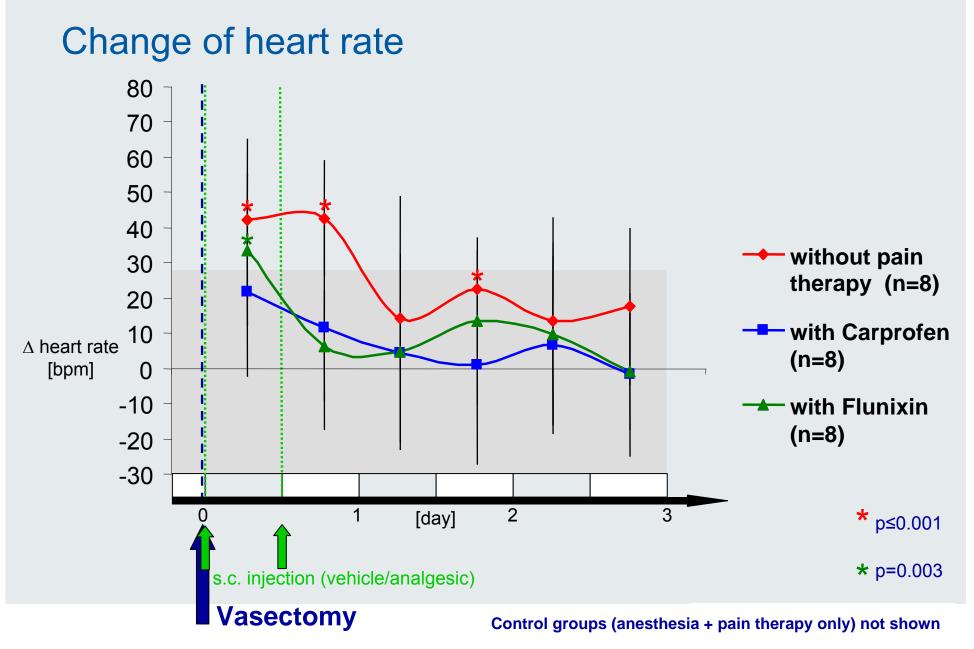


Recovery of mice after minor surgery with and without pain killers (NSAID: Carprofen, Flunixin)

Telemetric measurements \rightarrow preliminary implantation of transmitters

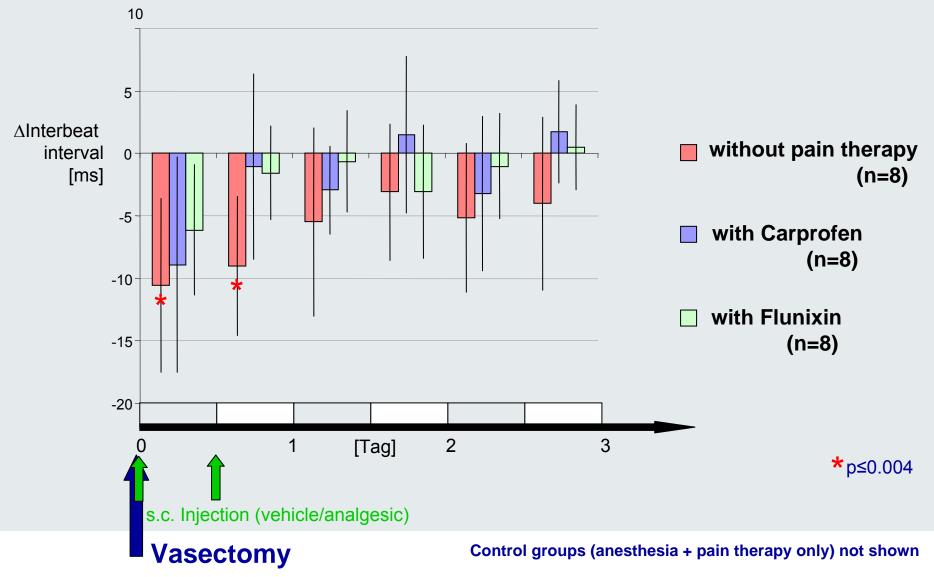






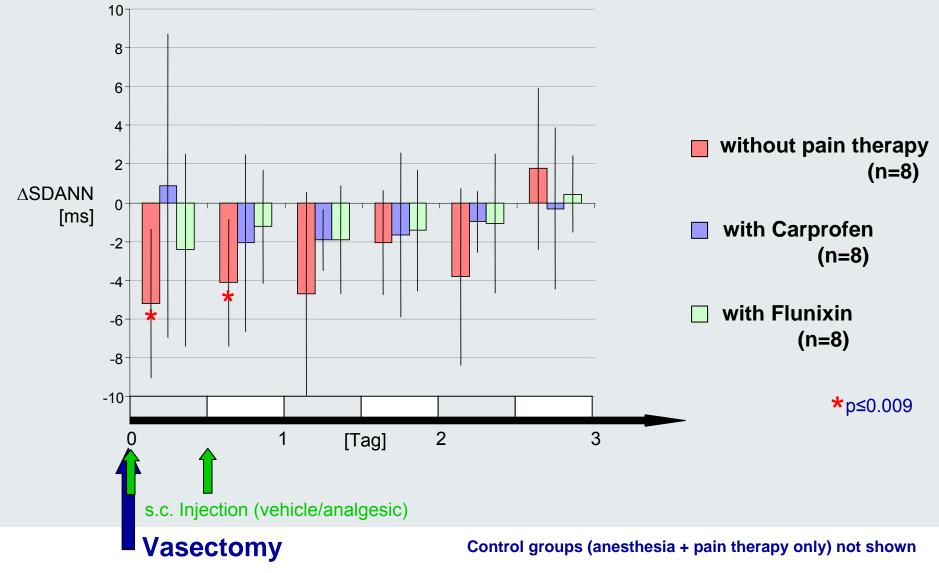
Arras, M. et al. BMC Vet Res; 3:16 (2007)

Time domain analysis of heart rate variability: IBI



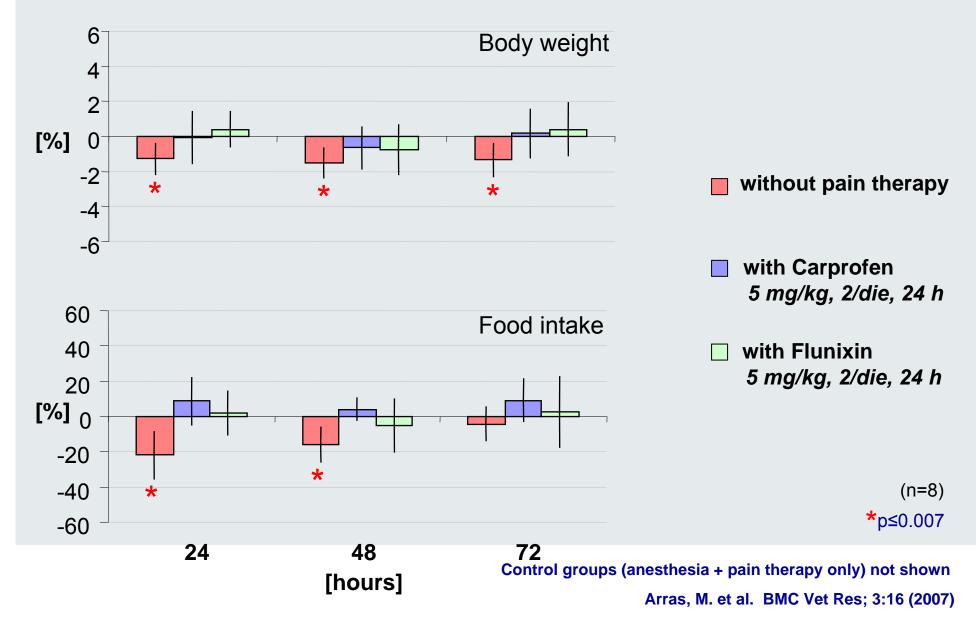
Arras, M. et al. BMC Vet Res; 3:16 (2007)

Time domain analysis of heart rate variability: SDANN



Arras, M. et al. BMC Vet Res; 3:16 (2007)

Body weight and food consumption



Recovery of mice after minor surgery with and without pain killers (NSAID: Carprofen, Flunixin)

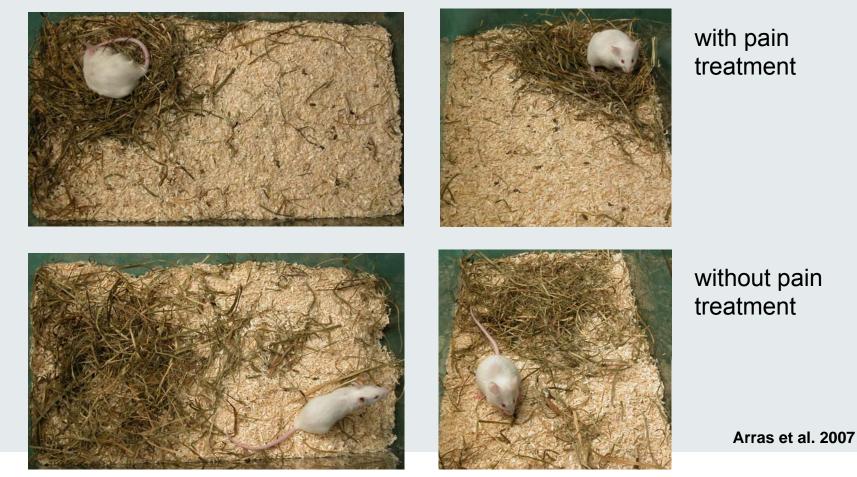
Attenuation of aberrations in heart rate and heart rate variability with analgesia Inhibition of the reduction in body weight (~2%) and food consumption (~20%) with analgesia

Clinical investigation: no difference in outer appearance, posture, spontaneous behaviors

Arras et al. 2007



Recovery of mice after minor surgery with and without pain killers (NSAID: Carprofen, Flunixin)

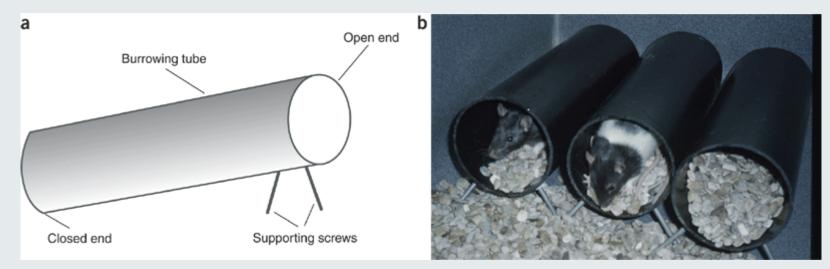




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Nature Protocols 1, - 118 - 121 (2006) Burrowing in rodents: a sensitive method for detecting behavioral dysfunction Robert M J Deacon

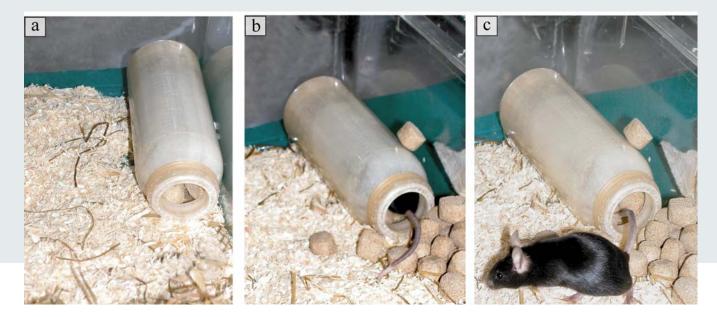


- > mice, rats, hamsters, gerbils
- home cage based
- > read out: aberration of natural, highly motivated behavior



Burrowing

Species-specific, spontaneously occurring behaviour High motivation to dig burrows persists during domestication Deacon et al. 2001 > neurodegenerative diseases Useful as indicator for impaired wellbeing???



Hypothesis: Abberration of a natural behavior is an indicator of pain, constraint, or impaired well-being

Aim: Changes in burrowing behavior as indicator of pain after minor surgery

Pre-study burrowing test

Time of testing (circadian rhythm), Setup (apparatus), Sex bias, Strain bias C57BL/6J, DBA/2J, 129Sv/Ev-IFNabRtmAgt

Study design

32 male & 32 female adult C57BL/6J



Burrowing test: study design

| | | limatisation g with burrowing a | pparatus | baseline measurement | | experimental measurement | |
|----|------------------------------|------------------------------------|------------|---------------------------|---------------------|---|---------------|
| | n = 16 | | | | | surgery + anaesthesia | |
| | | n = 16 | | | | surgery + anaesthesia + analgesia | |
| | | n = 16 | | | | anaesthesia | |
| | | n = 16 | | | | anaesthesia + analgesia | |
| | | | | | | | L |
| da | y 0 day 1 | day 2 | day | y 3 day | y 4 day | y 5 day | y 6 |
| | | | | video recording | | video recording | |
| | UniversitätsSpital Zürich | | Welfare of | Laboratory Animals – Role | of Refinement? Berl | Jirko in 13 Dec 2011 24 | f et al. 2010 |

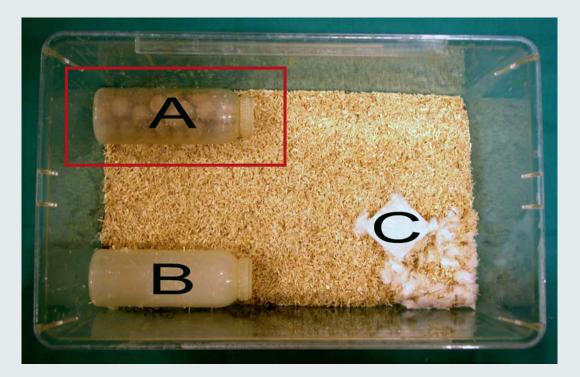
Burrowing test: study design

Surgical intervention

- Injection of analgesia (+/- carprofen, 5 mg/kg BW, s.c.)
- transport (5 min)
- inhalation anaesthesia (sevoflurane, 15 min)
- +/- one side minor laparotomy
- recovery on warming mat (at least 15 min)



Burrowing test: set up

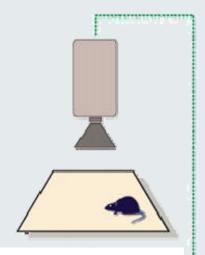


Experimental Setup. Burrowing test apparatus (A), shelter (B) and nesting material (C) in home cage.

Jirkof et al. 2010



Burrowing test: analysis



Before & after treatment Video recording Analyses: software based, ObserverXT (Noldus) Burrowing parameter:

- latency to burrow
- duration of burrowing
- weights of removed pellets after 2h





Results: burrowing performance

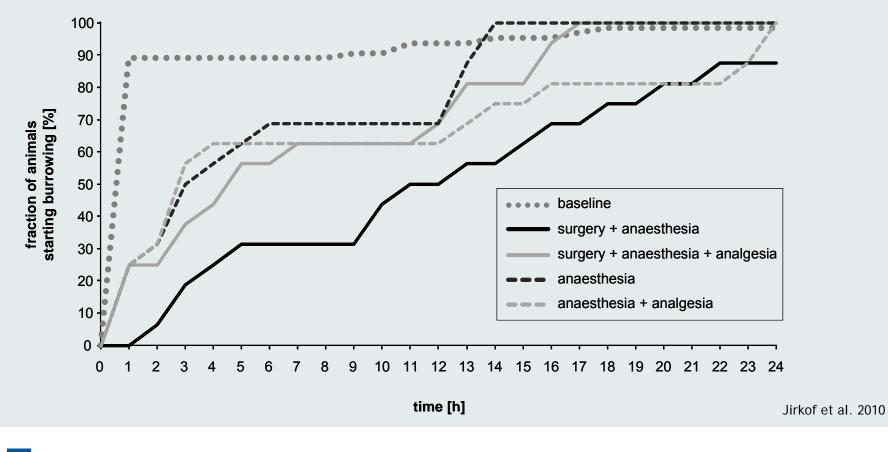
Almost all (98 %) healthy mice burrowed during baseline measurement

Example: infrared video recording of burrowing.

All parameters (latency to burrow, duration of burrowing, weight of removed pellets) responded to the different experimental treatments.



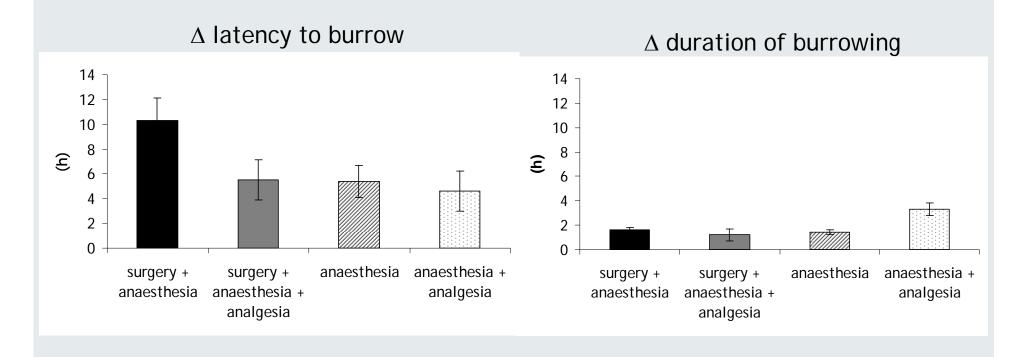
Results: burrowing performance





Results: baseline vs experimental

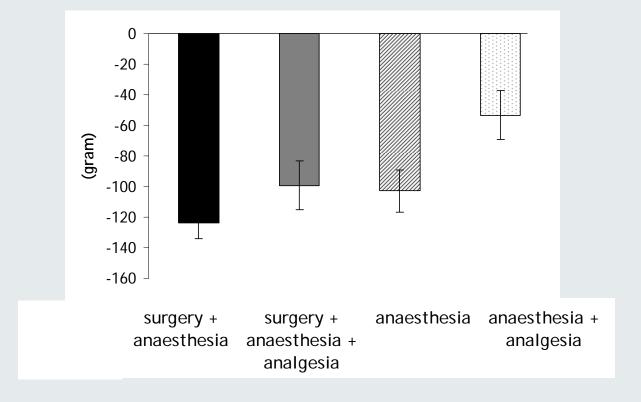
Increase in latency & duration of burrowing after treatment





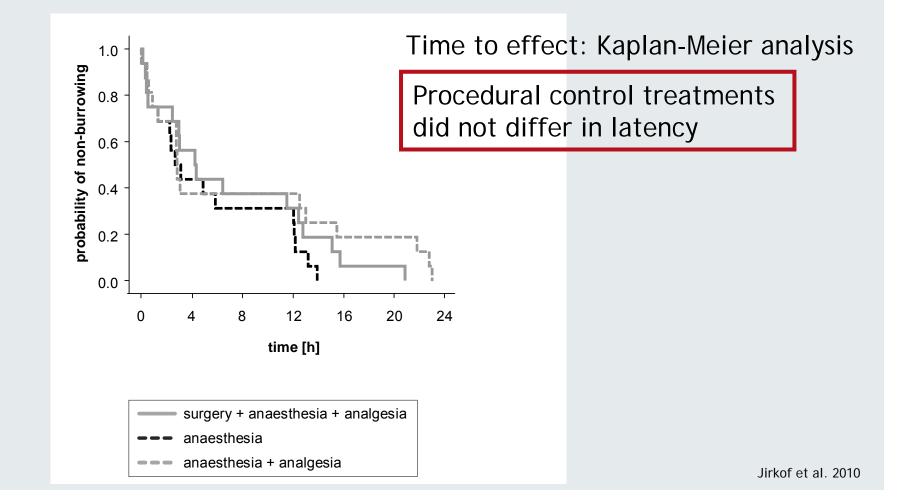
Results: baseline vs experimental

 Δ weight of removed pellets after 2h



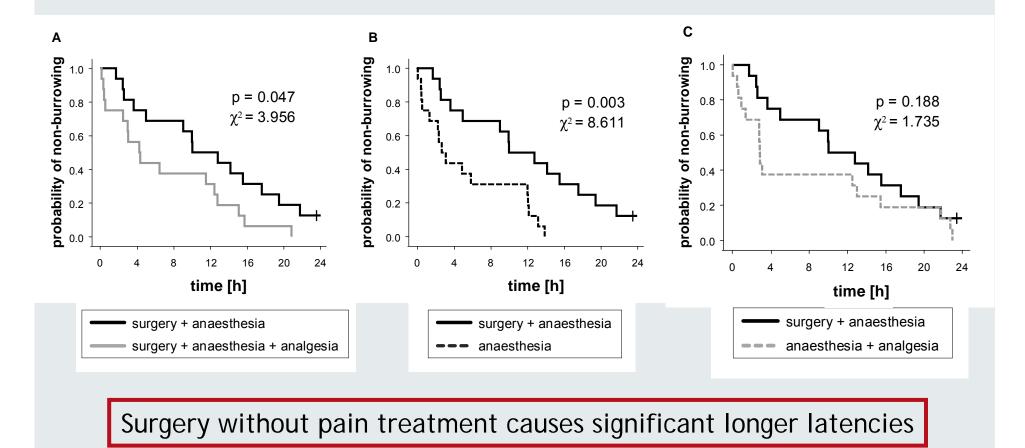


Results: group comparisons





Results: group comparisons



Jirkof et al. 2010



Burrowing test: summary & conclusion

- Easy to perform in the animals home cage under routine laboratory conditions
- Burrowing parameters react sensitive on treatments
- Results allow to discriminate between groups of mice that are treated with a pain killer or not before surgery
- Results imply a balance between the motivation to burrow and an individuals level of constraint experienced

In conclusion, postsurgical impairment, mainly attributable to pain, can be assessed on the basis of the burrowing test



Where are we now?

Currently: new approaches and methodologies for the assessment of pain and constraint in mice

- Conditioned place preference → Frank Porreca [King et al. 2009]
- Facial expression (mouse grimace scale) → Jeff Mogil [Langford et al. 2010]
- General changes in behaviors (rearing, climbing, etc.) \rightarrow van Loo et al. 2007, Roughan et al. 2009
- Abberrant behaviors (e.g. twitching, flinching, ...) → Paul Flecknell [Roughan et al. 2009]
- Drug self administration → Vera Baumans [Pham et al. 2010]
- Alterations of natural behaviors → Jirkof et al. 2010
- Sympathetic reaction (telemetry) → Arras et al. 2007
- Ultrasound vocalisations?

Feasibility, validity, benefits, limitations in routine laboratory conditions Reliability of specific tests for estimating the efficacy of pain relief regimens



Long-term goals

Utilization of new tests for verifying pain relief regimens which are in use to-date

Establish efficient, evidence-based modern pain relief regimens

- Sustained-release drugs
 - Foley et al. 2011
- Administration of analgesia via drinking water, food, or mini-pumps
- Multimodal analgesia protocols
- ...

Is there a market for analgesics or other drugs for use in mice???



Thank you for your attention

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